Project No. __________
Book No. __________

Objectives: Prepare 9 x 700 ml of XLD. (3/6/07)

Procedure:
1) Use cylinder to measure 700 ml of DI water and pour it into the flask.
2) Disregard the line of the water and discard some amount of water.
3) Place 38.5 g of XLD sugar (25 g/l) and carefully pour it into the flask.
4) Pour DI water to fill 700 ml.
5) Drop stir bar and cover with foil, label.
6) Heat mix until media boils.
7) Cool down on the water bath (50°C) 40 mins.
8) Pour plates.

Do housekeeping for the lab 3.15.
Continue with the inventory check of the equipment.
Objective: To prepare 1 x 700 ml TSA, 1 x 700 ml MM, 2 x 9 ml ± 2 ml of BPN agar

Procedures:

I. Prepare 1 x 100 ml of TSA
1) Use cylinder to measure 900 ml DI water and mark the water line
2) Discard some water
3) Weight 25 g of TSA and pour into the flask (1 L = 419 g)
4) Add water to the marked line
5) Heat mix using stir bar while covering with foil
6) Autoclave at 121°C 45 mins
7) Cool down in water bath (50°C) for 40 mins
8) Pour plate using aseptic technique

II. Prepare 1 x 100 ml MM
1) Mark the flask at the volume of 100 ml using DI water and cylinder to measure the amount of water
2) Discard some water
3) Weight 40.25 g of oxoid medium base and pour it into a flask
   (37.5 g/l, 0.7 L = 40.25 g)
4) Add DI water to 70 ml
5) Autoclave at 121°C 45 mins
6) Cool down in water bath for 40 mins (50°C)
7) Add 5 ml of modified mofford medium supplement
8) Mix 30 sec.
9) Pour plate

III. Prepare 2 racks of BPN agar
1) Calculate the amount of medium needed to prepare (9.5 x 75 x 2) + 500 = 15 L
2) Use cylinder to measure 1.5 L DI water, mark the line
3) Discard some water
4) Weight 80 g of BPN media (209 g/L) into 1.5 L you need 30 g
5) Deep in magnetic bar and heat mix while covering the flask with foil
6) Cool down on bench
7) Distribute into tubes using dispensers
8) Autoclave at 121°C 45 mins
9) Cool down on bench
   DPD - Collect the hazard disposal

Witnessed & Understood by me:

Date: 3/12/07
Signed: [Signature]
Invented By: [Signature]
Date: 3/2/07
Recorded by: [Signature]
Objective: To prepare 750 ml of XLD.

Procedure:
1. Use cylinder to measure 700 ml of DI water. Use label to mark the line.
2. Discard some amount of water.
3. Weigh 30.8 g of XLD media, and carefully pour into the flask (55.946)
4. Turn on the water bath.
5. Heat mix 9:10 until the media boils.
6. Cool down in a water bath. 40 mins (50°C).
7. Pour plate.

- Do the housekeeping for labs 314, 315, 305.
- Do hill run.
Objective: To prepare 2 x 700 ml TSB and 2 x 700 ml M0X

Procedure:
1. Use cylinder to measure 700 ml of DI water.
2. Mark the water line.
3. Pour out some water.
4. Weigh 25 g of TSB media (50 g/l) using weight boat.
5. Carefully pour the media into the flask, add DI water to 700 ml.
6. Drop stir bar and cover with foil.
7. Heat mix
8. Auto close, 1 hr 45 min.
9. Cool down in a water bath for 40 mins (90°C).
10. Pour plate

For 2 x 700 M0X
1. Use cylinder to measure 700 ml of DI water, use marker to mark the volume.
2. Discard some water.
3. Weigh 40.25 g of media on a weight boat (525 g/l).
4. Add DI water to 700 ml.
5. Auto close, 1 hr 18 min 45 mins.
6. Cool down in water bath for 40 mins (80°C).
7. Add 1 ml of modified Oxford medium supplement in to the agar flask.
8. Mix for 20 sec.
9. Pour plate

Do the housekeeping for lab 3/4/07, 3/5/07.
- Collect hazard labels
Objective: To prepare 9 ml of buffer phosphate buffer

1. Calculate the amount of media that is needed to prepare (4x7.5 - 9.5x7.5 x 4) 3.2 L of PBS buffer chilled at 4°C.
2. Use cylinder to measure 3.2 L glassware and mark where the water lies.
3. Discard some amount of water.
4. Weigh and add 1.64 g of PBS (20 g/L).
5. Carefully pour the media into the flask.
6. Add DI water to 3.2 L.
7. Heat mix until the media dissolves. - buffers don't need to be heated.
8. Distribute into tubes using dispenser.
9. autoclave at 121°C for 45 min.
10. Cool down on bench.

- Do housekeeping room 44
- Do hill run
- Do inventory check of the expired media.