OBJECTIVE: To prepare 10 x 700 ml BHI.

1) Make the 1 L flask by using cylinder and 3 l water to mask the volume at 700 ml.
2) Discard some amount of water.
3) Weigh 36.4 g of BHI.
4) Carefully pour the agar into the flask.
5) Fill the water into the volume masked.
6) Drop in magnetic bar and cover with foil. label with an autoclave tape.
7) Heat mix.
8) Pour some amount into a basket to measure pH.
9) Autoclave 45 min 121° C.
10) Put it in 50°C water-bath (45°C) for 10 mins.
11) Mix 20 s.
12) Pour plate.
Objectives: To prepare 2 x 700 ml of BHA

1) Use 100 ml cylinder to measure 700 ml of DI water
2) Make each flask 3/4 full at the level of 700 ml. Discard some water
3) Weigh 26.4 g of BHA C 1g use 52.9 g
4) Carefully pour the media in to the flask
5) Heat mix and place under cover with foil and heat mix
6) Autoclave 121°C 45 min
7) Cool in water bath (25°C) for 40 min.
8) Mix 20 sec.
9) Pour plate.

Do house cleaning for each lab.
- Do housekeeping 4.30, 3/4, 3/17, 30/5

1. Cool in water bath (40°C) 40 min.
2. Add 
  1.2 g/75 ml
3. Heat mix
4. Add
  5. Pipette 10 µl of 20% RNAse and 10 µl of 0.1M
6. pipet against 10 ml COPE and cover with plastic foil.
7. Place in the block to room
8. and 
9. Place in the room at 80 or 85°C
10. and then fill into the glass
11. with the point of 40°C and by using syringe and 0.5 µl
12. Place in 80°C for 15 min
13. Cool on bench.
14. Add
15. Buffer 1°C, 0.5 M
16. and
17. Add 5 µl of the sample
18. The solution to measure 0.5 ml from the 300 µl and use it to measure 96 µl.
19. Since 125 µl
20. is 4.15 ml
21. and
22. Place the micropipette, sterile solution
23. (10 µl 1.4 ml + 300 µl 25°C, 4°C)
24. Solution: 25% solution of buffer B, pool 10 ml
25. Project No.: 00-180, 00-180, 00-180
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Objective:
1. Prepare 2 x 100 ml TSB
2. Prepare 2 x 100 ml MAX
3. Prepare 4 racks 9.5 ml of Buffered peptone water.

Procedures:
1. Prepare 2 x 100 ml TSB.
2. Use cylinder to weight DI water 100 ml and pour the water into the flask.
3. Make the plate where the media lies.
4. Discard some water.
5. Use weight boat to weight 25 g of TSB media (1L use 40 0.7L use 25)
6. Carefully pour the media into the flask.
7. Pour DI water to the marked line.
8. Drop a stir bar and cover with foil.
10. Pour some media into 10 for pH measurement.
11. Sterilize by autoclaving at 121°C, 15 min.
12. Cool down in water bath (48°C) for 40 mins.
13. Pour plate.

I. Prepare 2 x 100 ml MAX (constituent medium base)
1. Use cylinder to weight DI water 100 ml and pour the water into the flask.
2. Make the water line and discard some water.
3. Weight 40.29 g of media on a weight boat (41.59/1L, 0.7 L = 40.25)
4. Drop in a magnetic bar, cover with foil, heat mix.
5. Sterilize by autoclaving at 121 °C 15 min.
6. Cool down in water bath (48°C).
7. Prepare modified 10% medium supplement by rehydrate the supplement by adding 10 ml of DI water.
8. Add supplement to the 300 media after the media cool down (121°C) 48 min.
9. Pour plate.
10. Leave to cool down on the bench.

IV. Prepare 4 racks of Buffered peptone water (9 ml)
1. Calculate the amount of media (9.5 x 9 x 75) = 660 : 8.5 L.
2. Mixed 2 x L into line by using cylinder and DI water.
3. Weight 64 g of BPN (2.64 g/L of water)
4) heat mix 85.69
5) Distribute into tubes by using dispenser
6) Autoclave 121°C, 15 min.
7) huge on a bench to cool down
8) Do daily housekeeping in lab 315, 315
9) Do weekly housekeeping in lab 314, 315, 305
10) Do inventory check for lab 314, 315 equipment.
Objective: To prepare 9 x 700 ml of TSA. (3.14 x 2)
90 prepare 6 x 1 L of 1x PBS and 6 x 500 ml of 1x PBS

I) Prepare 9 x 700 ml of TSA
   1) Use cylinder to measure 700 ml of DI water and pour it into 1 L flask.
   2) Mark the line of water and pour out some water.
   3) Weigh 28 g of media using weighboat and balance. (to 9/4)
   4) Carefully pour the TSA into the flask. Add water to 700 ml.
   5) Heat mix using stir bar, also cover with foil.
   6) Autoclave 210°C for 30 min.
   7) Cool down in water bath (40°C) for 40 min
   8) Pour plate

II) 1x PBS (to 1 L and 500 ml)
   1) Add 1 L of DI water to 1 x PBS measure 900 ml of DI water and autoclave 121°C, 45 min
   2) Add 100 ml of sterile 1x PBS into 900 ml sterile water to make 1x PBS 1.0 M
   3) Add 500 ml of sterile 1x PBS into 450 ml sterile water to make 1x PBS 0.5 M
   4) Cap tightly and shake vigorously to thoroughly mix it.

Witnessed & Understood by me: [Signature]
Date: 3/7/07
Invented by: [Signature]
Date: 3/8/07
Recorded by: [Signature]